

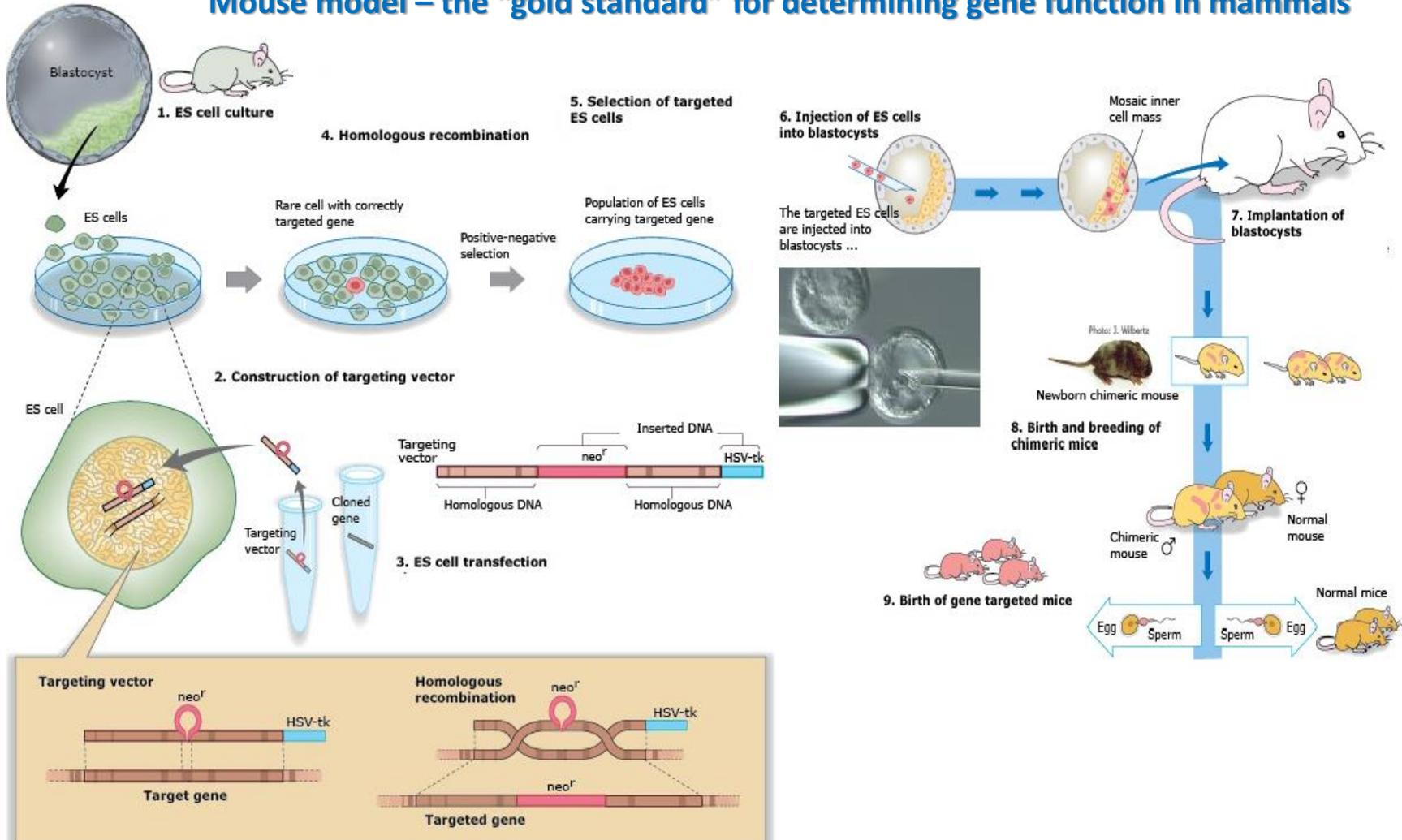
Talens and CRISPR/Cas9: Generation of genome engineered mouse models using editable nucleases

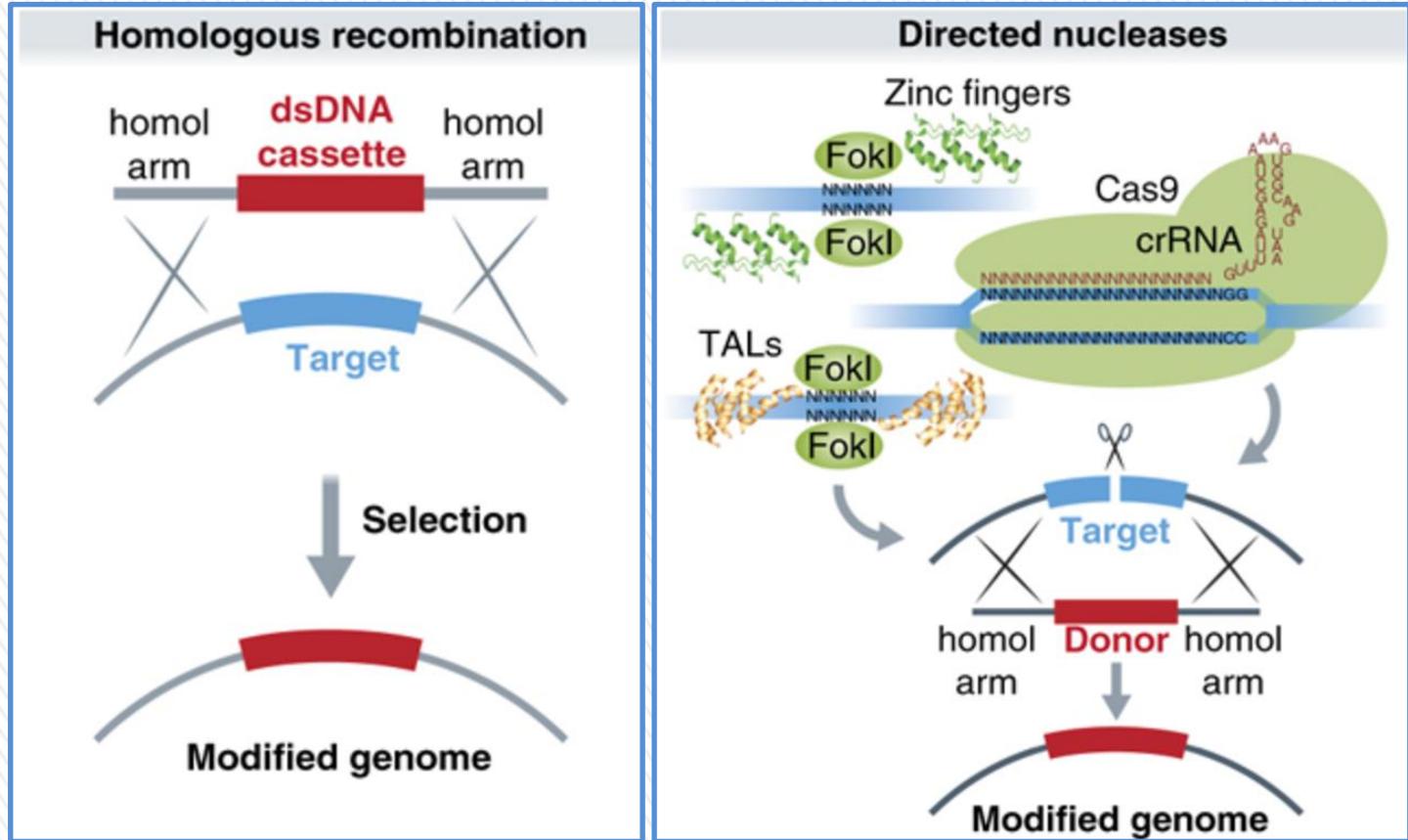
Silvia Petrezsélyová, PhD & Dominika Fričová, PhD



Institute of Molecular Genetics of the ASCR, v.v.i.

Mouse model – the “gold standard” for determining gene function in mammals

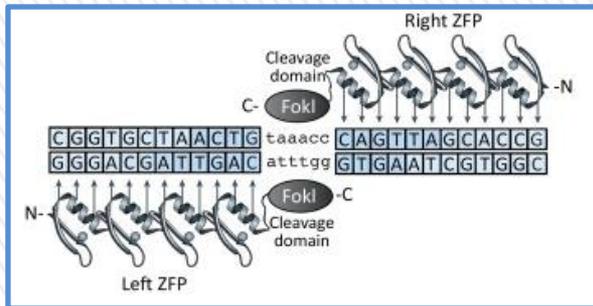




Induction of DSB increases HDR 100 – 1000 x

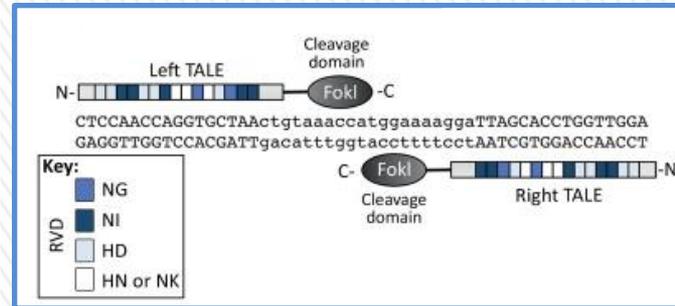
Esvelt and Wang, *Mol Sys Biol* 2013

Zinc Finger Nucleases



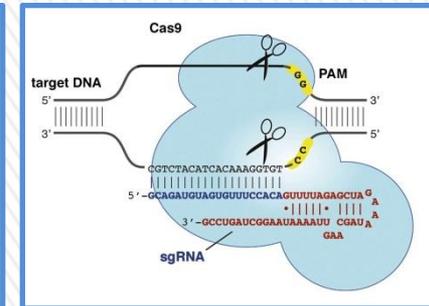
- Cys2-His2 zinc finger domain
- artificial arrays of 3-6 Zinc Fingers (9-18 bp)
- C-terminal fusion with endonuclease (FokI) - ZFN

Transcription Activator-like Effectors Nucleases (TALENs)



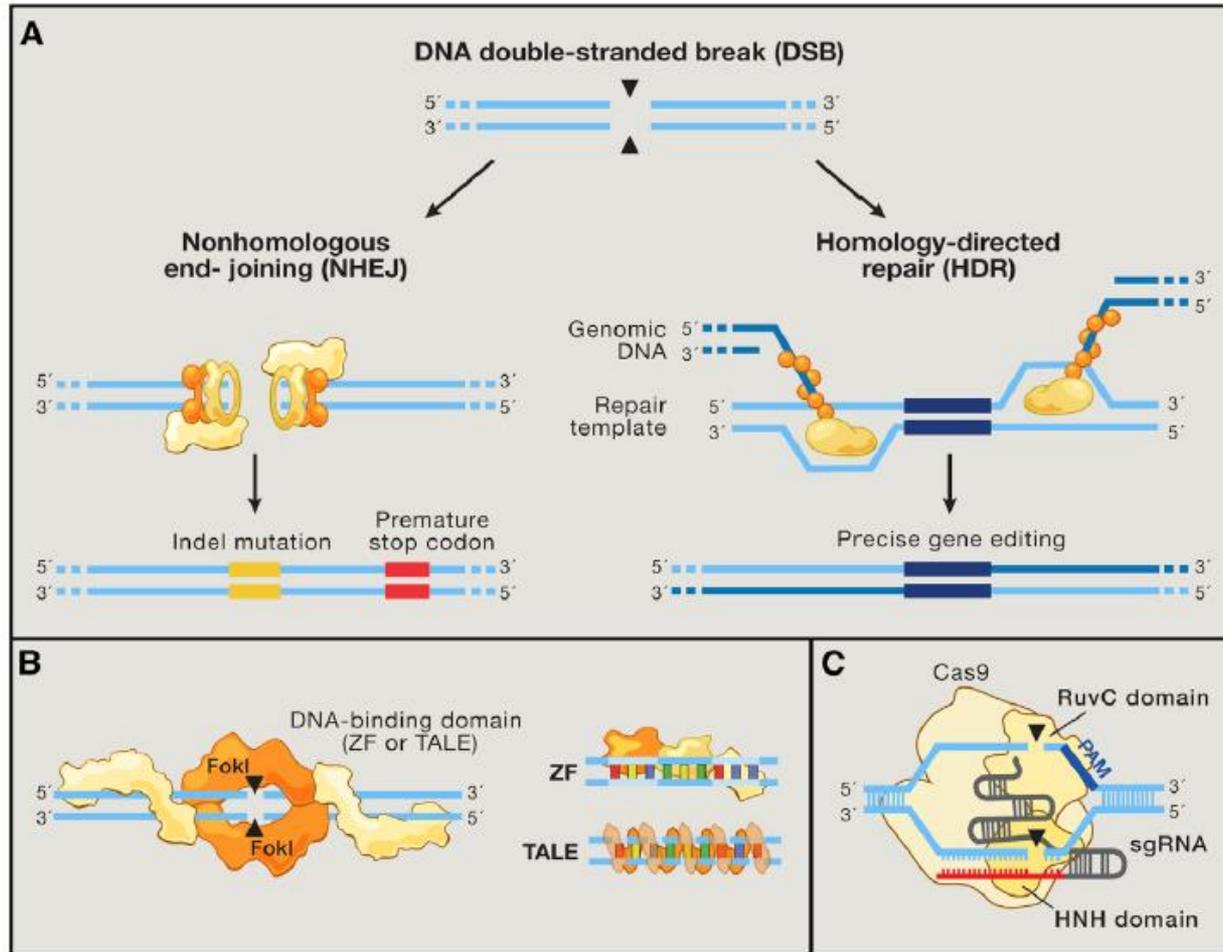
- Central Repeat Domain (CRD) responsible for DNA binding
- CRD consists of 34 aa highly homologous repeat modules
- DNA specificity determined by aminoacids 12 and 13 of each repeat - repeat variable diresidues (RVDs)

CRISPR/Cas9 system



- Interspaced short palindromic repeats (CRISPR) systems
- CRISPR RNAs (sgRNAs) in complex with CRISPR-associated 9 (Cas9) protein

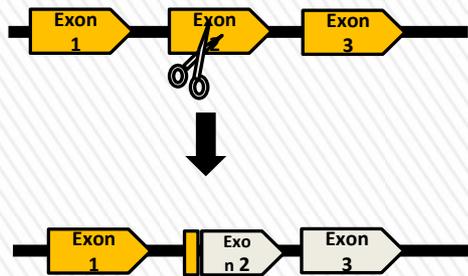
Gaj *et al.* (2013) Trends in Biotechnology
Jinek *et al.* (2013) Elife



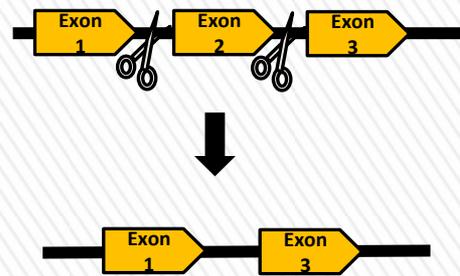
Hsu *et al.*, Cell 2014

possibilities of using programmable nucleases

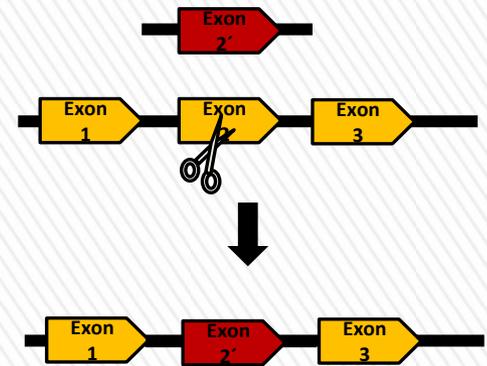
Generation of indel mutations



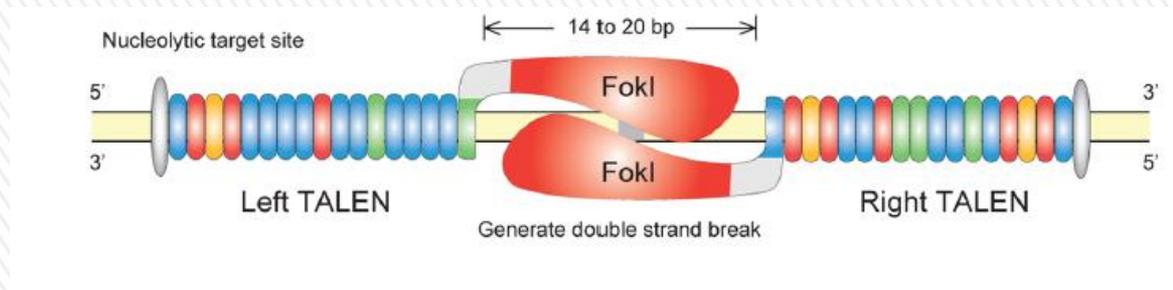
Excision of DNA fragment



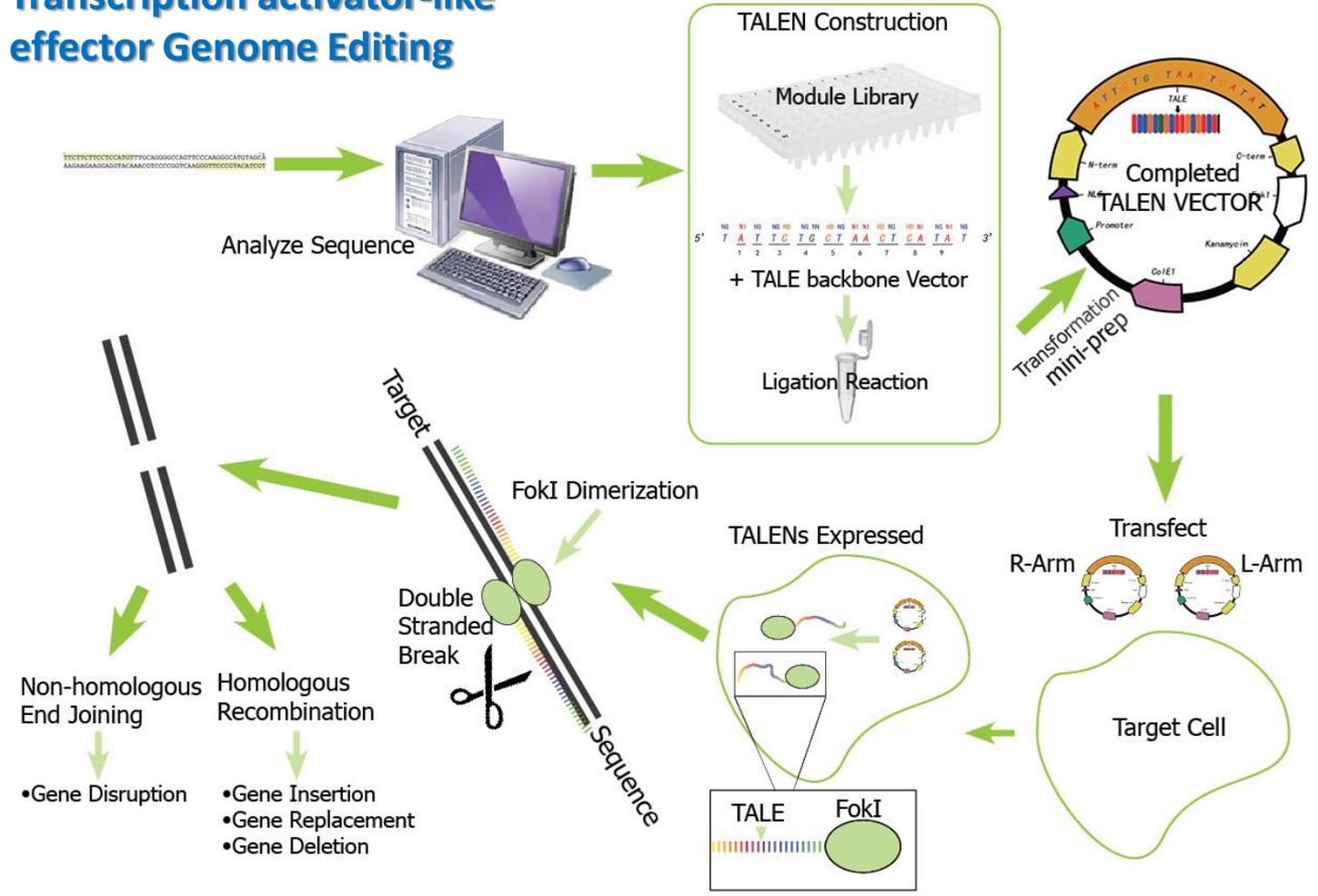
Site-specific integration



Transcription Activator-like Effector Nucleases (TALENs)



Transcription activator-like effector Genome Editing



TALEN design guidelines

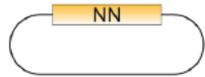
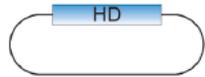
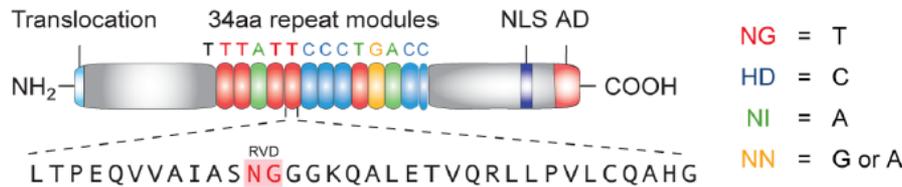
<https://tale-nt.cac.cornell.edu/node/add/talen>



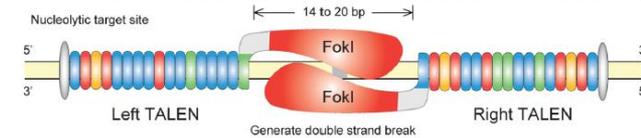
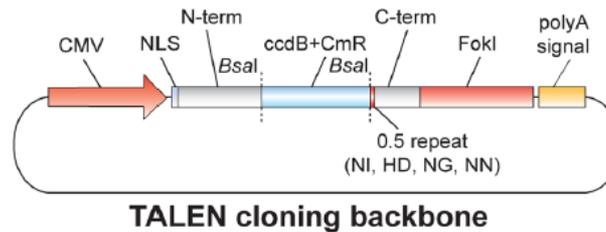
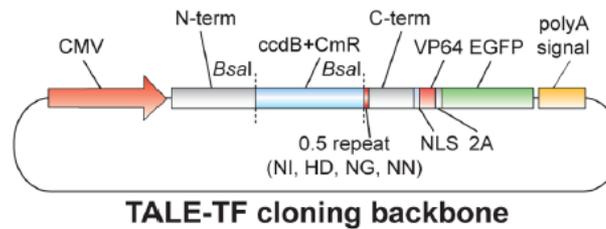
- TALEN target site should be preceded by 5' T
- TALEN should not have a T at position 1
- TALEN should not have an A at position 2
- Incorporate at least some (~3–4) strong RVDs (e.g., HD or NN)
- Use optimal spacer for your talen backbone (usually 15-17 bp)
- Position strong RVDs to avoid stretches (≥ 6) of weak RVDs
- For high guanine specificity use NH (to promote TALE activity) or NK
- Use NN for guanine if only a few other strong RVDs are present

Assembly of TALE nucleases

www.addgene.org Golden Gate TALEN and TAL Effector Kit 2.0 (Cermak *et al.*, 2011)



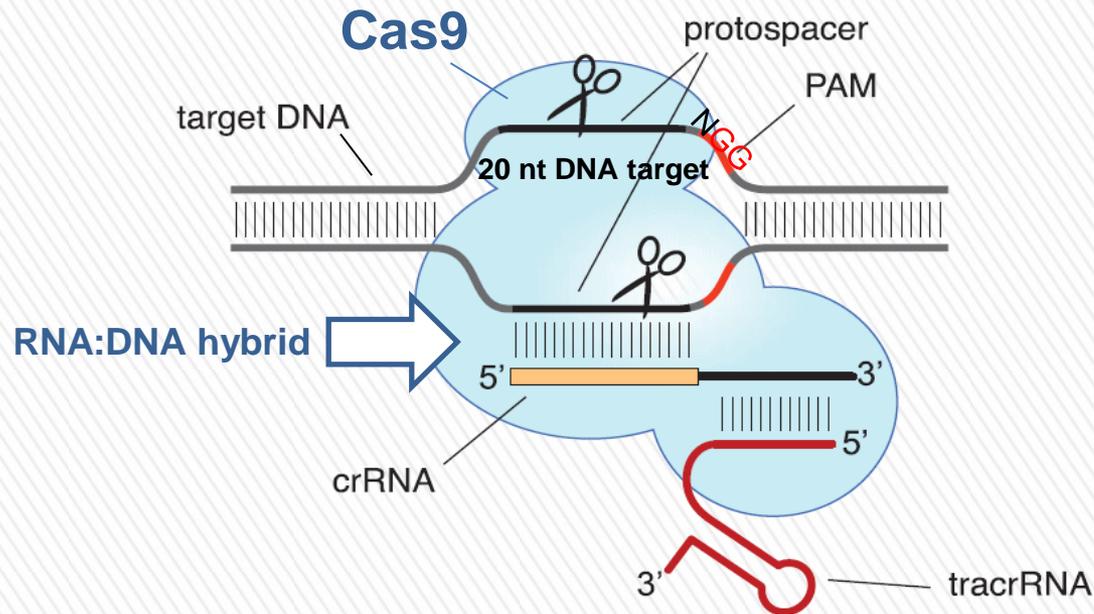
TALE monomer templates



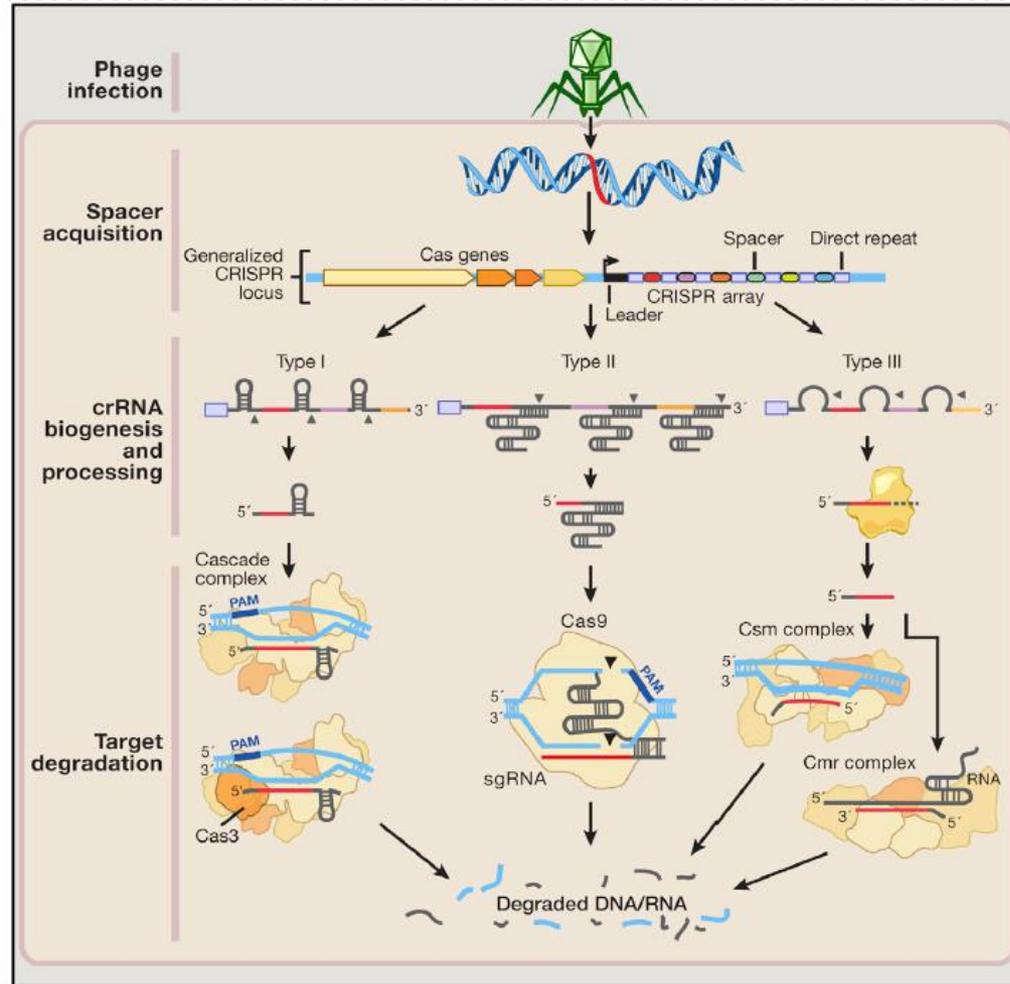
Sanjana *et al.*, Nat Protoc 2013

CRISPR/Cas9 Nuclease

CRISPR: Clustered Regularly Interspaced Short Palindromic Repeats
Cas: CRISPR-associated (protein)

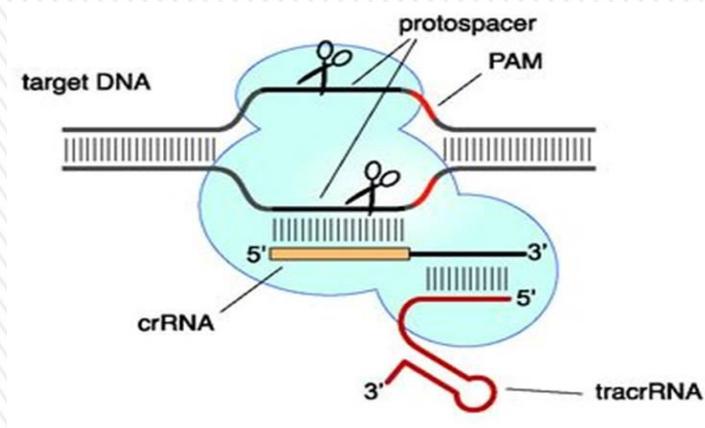


Natural mechanisms of Microbial CRISPR systems in Adaptive Immunity

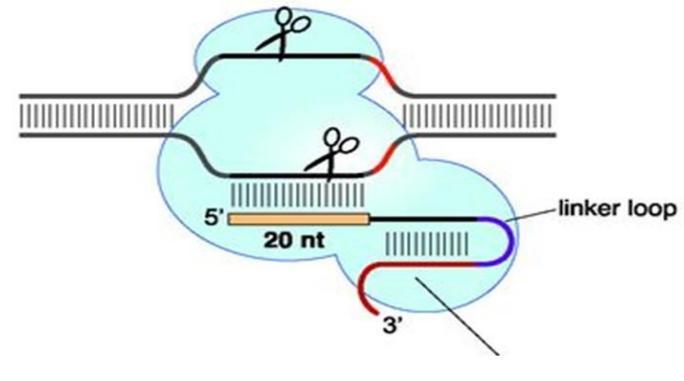


Hsu *et al.*, Cell 2014

Natural configuration Cas9 & crRNA:tracrRNA duplex



Simplified configuration Cas9 & single guiding (sg) RNA



Programming of Cas9 to a specific target sequence

Any 20 nt sequence, located upstream of a NGG (PAM) motif, can be addressed

- guides Cas9 to introduce a DSB 3 bp upstream of PAM

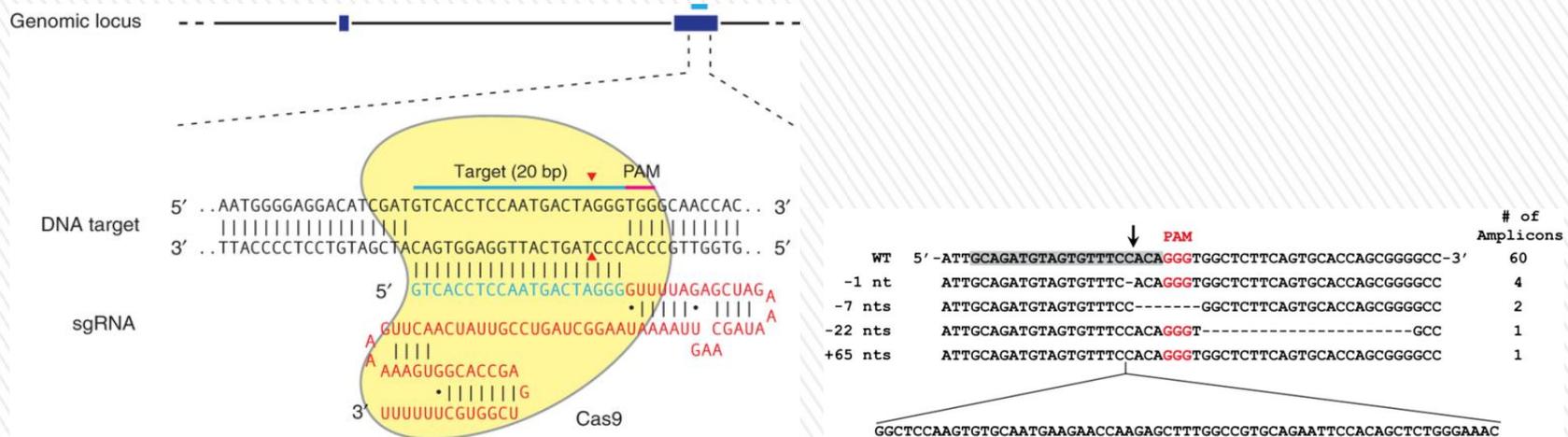
The Crispr/Cas9 system

<http://crispr.mit.edu/>

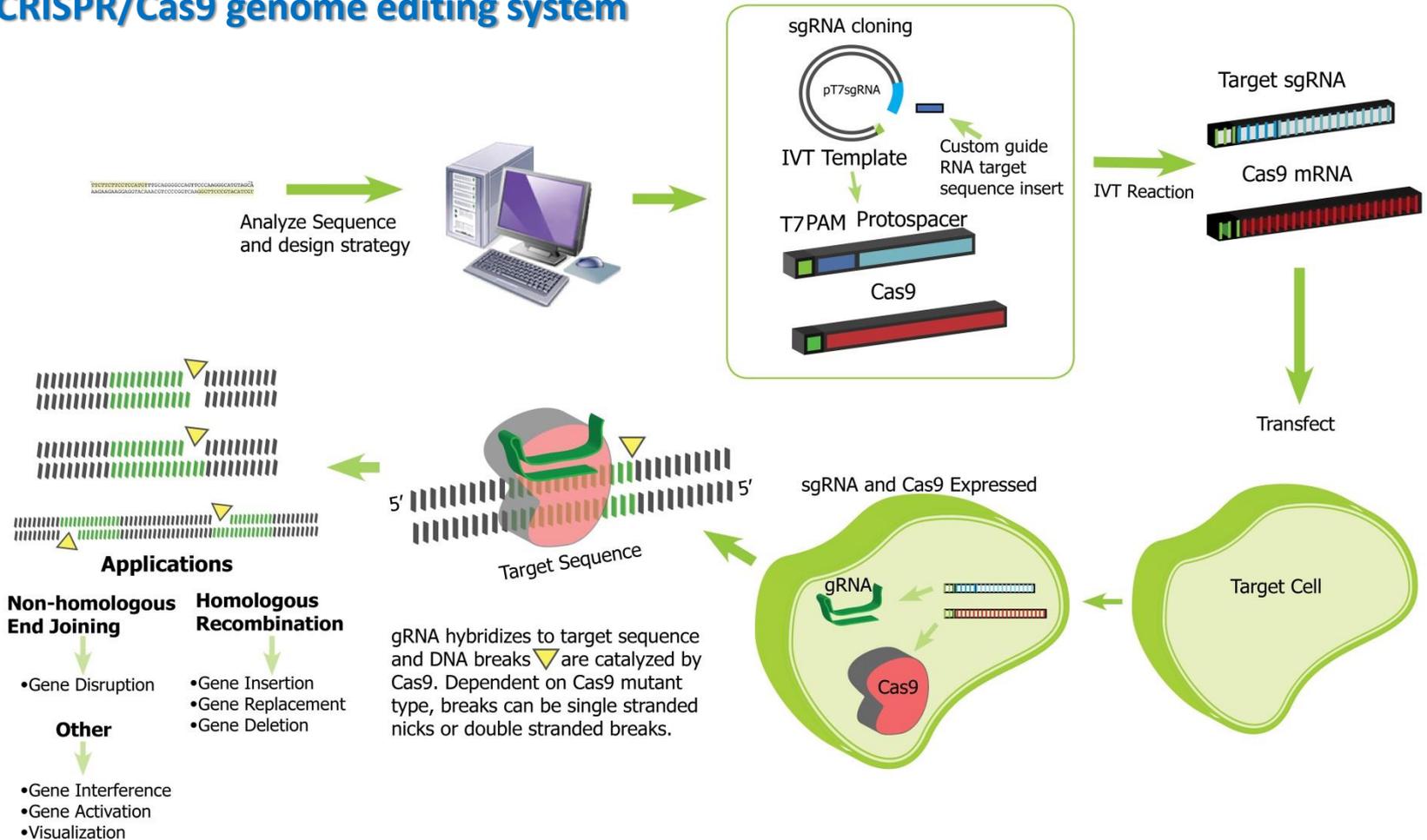
targeting

Cas9 nuclease is guided by small crRNAs to recognize and cut target DNA ("protospacer") sequences, next to an NGG motif (PAM – protospacer adjacent motif)

Functions efficiently for gene editing in mammalian cells (Mali et al., Cong et al., Science 2013).



CRISPR/Cas9 genome editing system



Generation of specific Crispr/Cas9:

Oligo annealing and cloning into backbone vectors:

easy task

1. Digest 1ug of pX260 or pX330 with *BbsI* for 30 min at 37C:

1 ug	pX260 or pX330
1 ul	FastDigest <i>BbsI</i> (Fermentas)
1 ul	FastAP (Fermentas)
2 ul	10X FastDigest Buffer
X ul	ddH ₂ O
20 ul	total

2. Gel purify digested pX260 or pX330 using QIAquick Gel Extraction Kit and elute in EB.

3. Phosphorylate and anneal each pair of oligos:

1 ul	oligo 1 (100μM)
1 ul	oligo 2 (100μM)
1 ul	10X T4 Ligation Buffer (NEB)
6.5 ul	ddH ₂ O
0.5 ul	T4 PNK (NEB)
10 ul	total

Anneal in a thermocycler using the following parameters:

37°C	30 min
95°C	5 min and then ramp down to 25°C at 5°C/min

4. Set up ligation reaction and incubate at room temperature for 10 min:

X ul	<i>BbsI</i> digested pX260 or pX330 from step 2 (50ng)
1 ul	phosphorylated and annealed oligo duplex from step 3 (1:250 dilution)
5 ul	2X Quickligation Buffer (NEB)
X ul	ddH ₂ O
10 ul	subtotal
1 ul	Quick Ligase (NEB)
11 ul	total

5. (optional but highly recommended) Treat ligation reaction with PlasmidSafe exonuclease to prevent unwanted recombination products:

11 ul	ligation reaction from step 4
1.5 ul	10X PlasmidSafe Buffer
1.5 ul	10mM ATP
1 ul	PlasmidSafe exonuclease
15 ul	total

Incubate reaction at 37C for 30 min.

6. Transformation

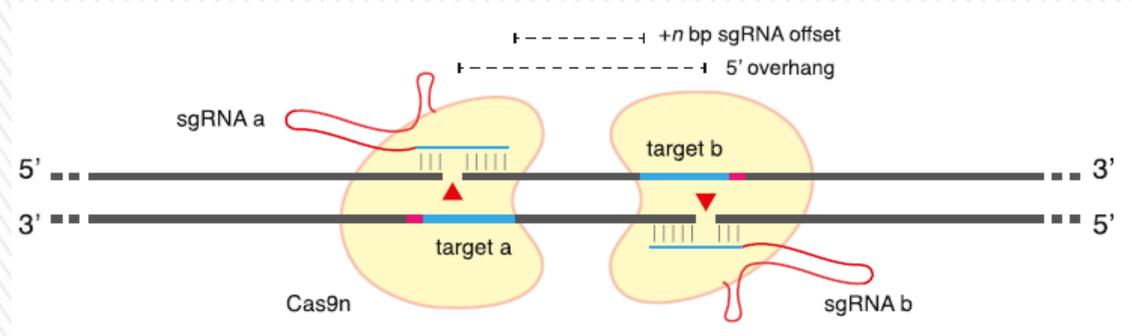
The Crispr/Cas9 system

increasing specificity

(low off-target activity)

Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity

F. Ann Ran,^{1,2,3,4,5,11} Patrick D. Hsu,^{1,2,3,4,5,11} Chie-Yu Lin,^{1,2,3,4,6} Jonathan S. Gootenberg,^{1,2,3,4} Silvana Konermann,^{1,2,3,4} Alexandro E. Trevino,¹ David A. Scott,^{1,2,3,4} Azusa Inoue,^{7,8,9,10} Shogo Matoba,^{7,8,9,10} Yi Zhang,^{7,8,9,10} and Feng Zhang^{1,2,3,4,*}



Offset from 4bp to 20bp

Programmable nucleases:

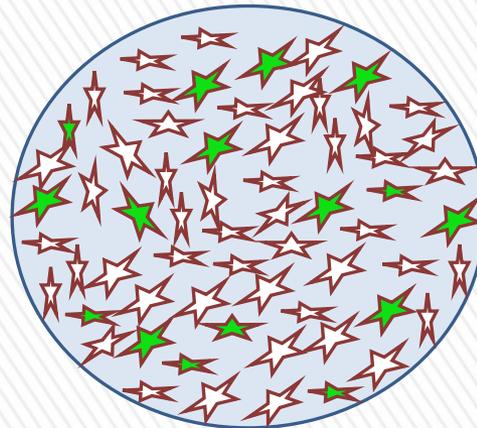
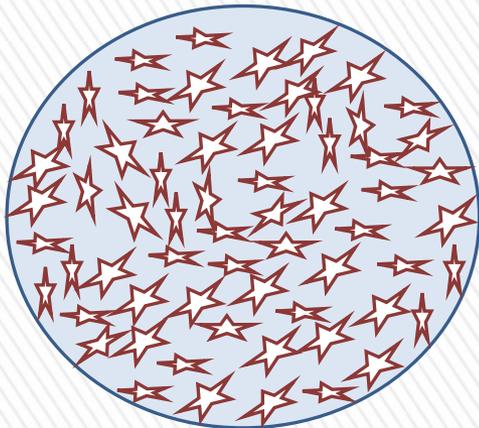
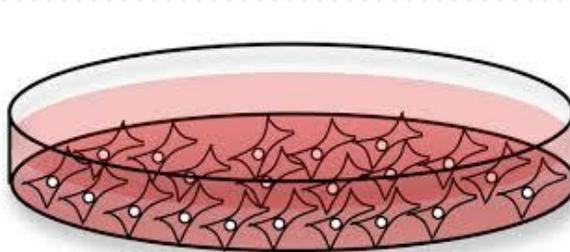
cell culture / *in vitro* validation

1) Design and generation of TALEN/CRISPR plasmids

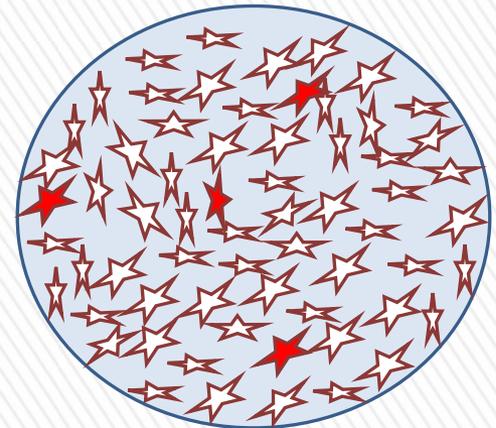
2) Transfection of cells

3) Selection of clones

TALEN/CRISPR plasmid DNA

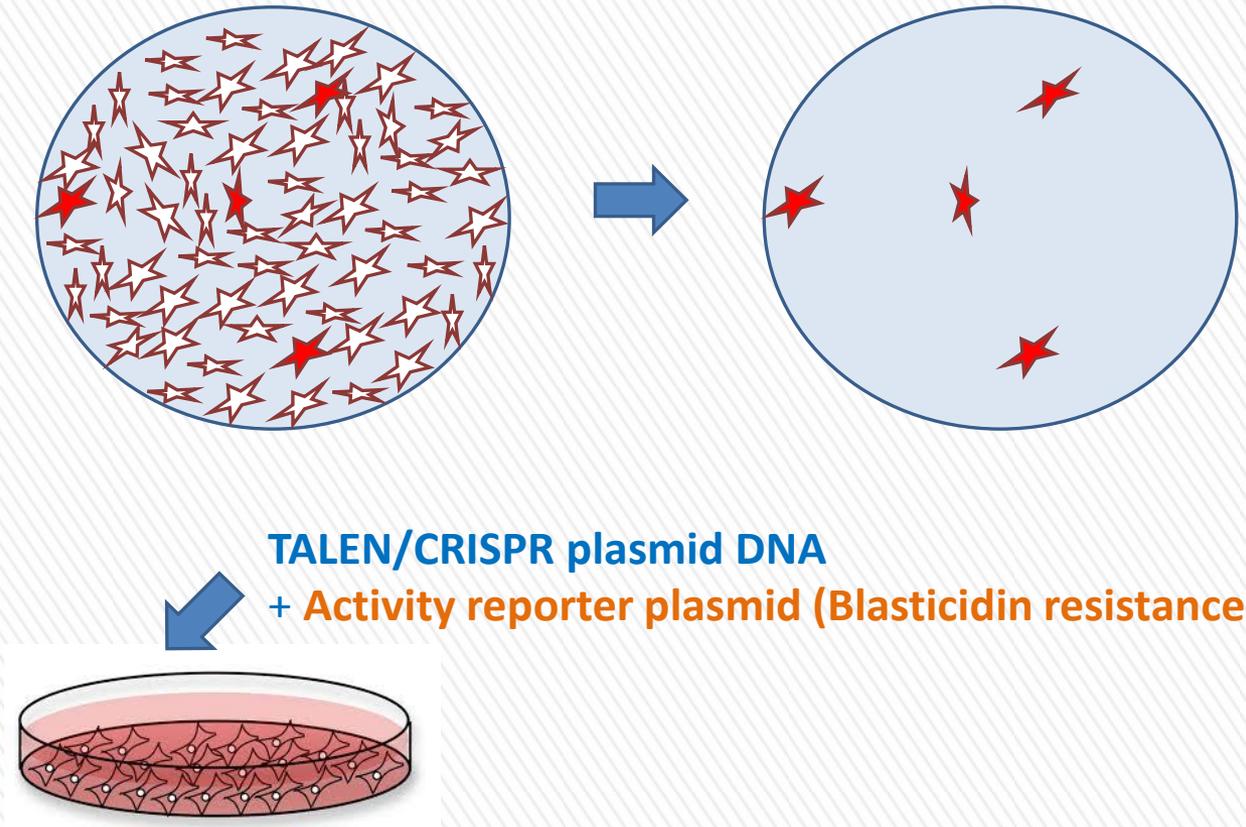


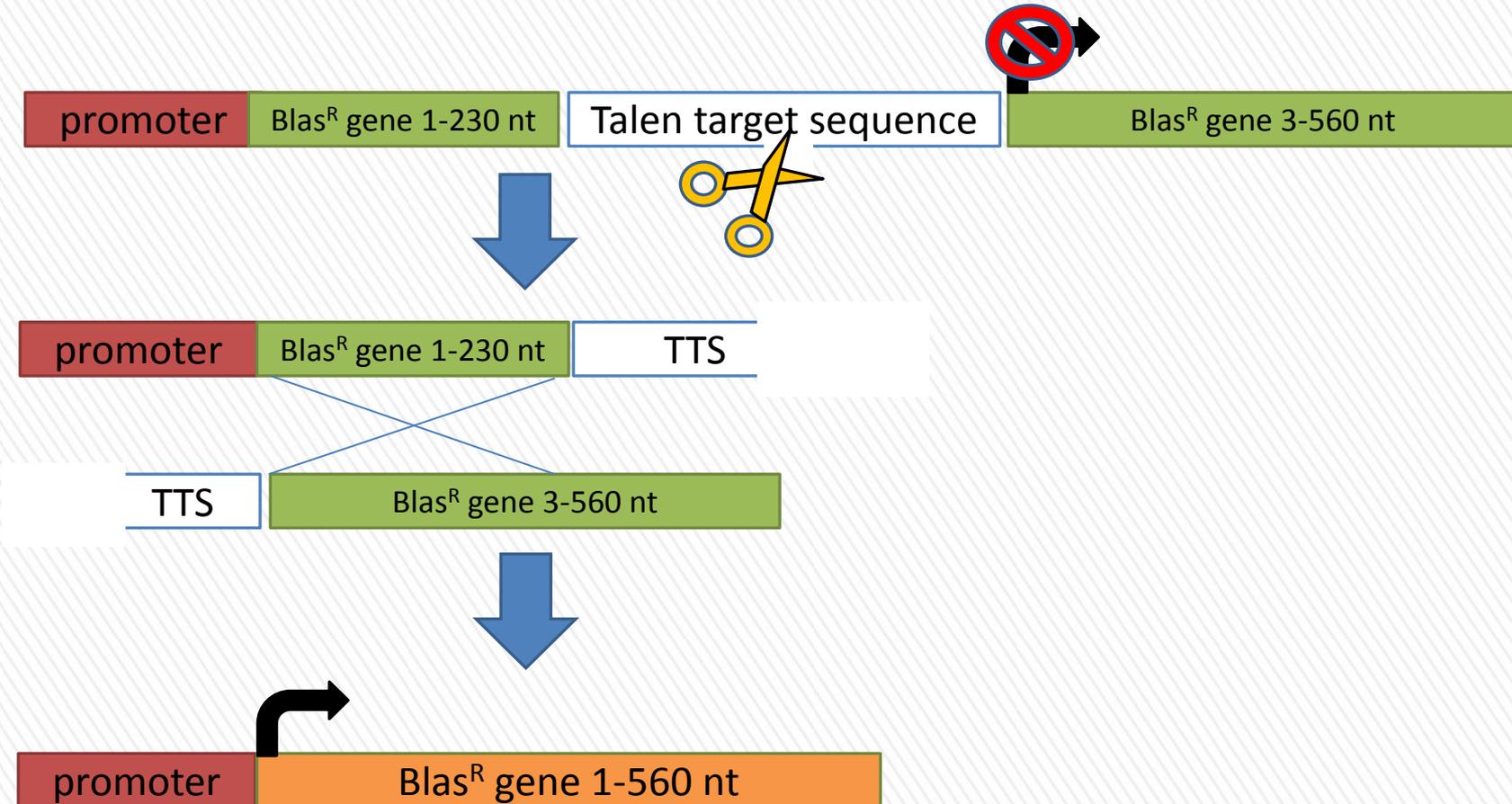
Transfected cells (15 %)



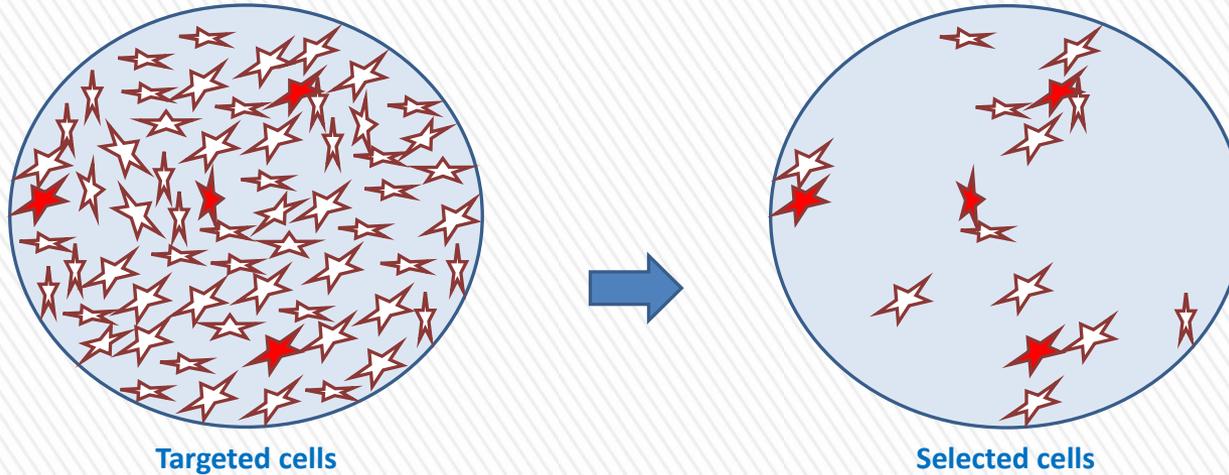
Targeted cells (1.5 %)

Surrogate plasmids for selection of targeted cells





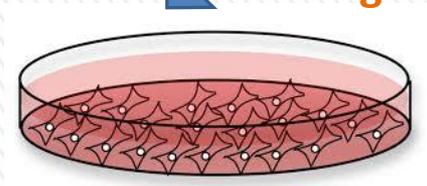
Kasperek- *unpublished data*



TALEN/CRISPR plasmid DNA

+ Activity reporter plasmid (Blasticidin resistance)

+ targeting construct

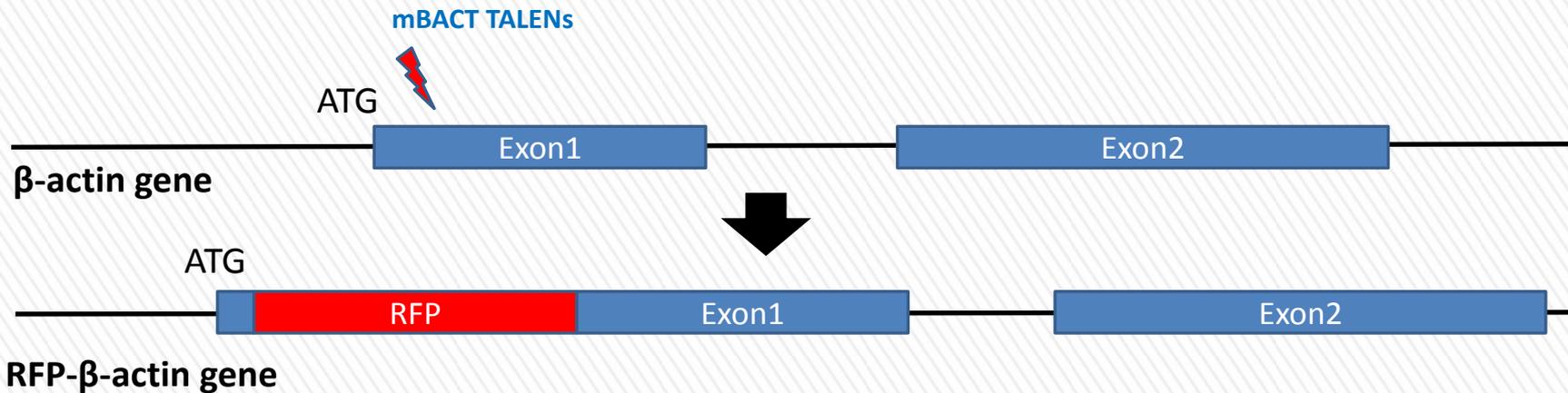
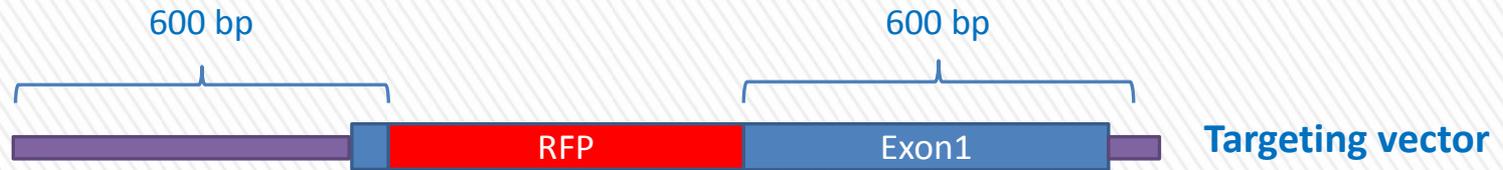
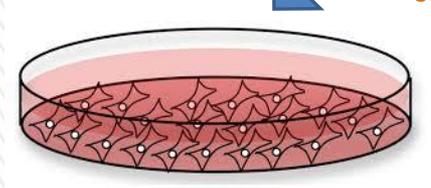


Kasperek- unpublished data

TALEN/CRISPR plasmid DNA

+ Activity reporter plasmid (Blasticidin resistance)

+ targeting construct



Kasperek- unpublished data

TALEN/CRISPR plasmid DNA

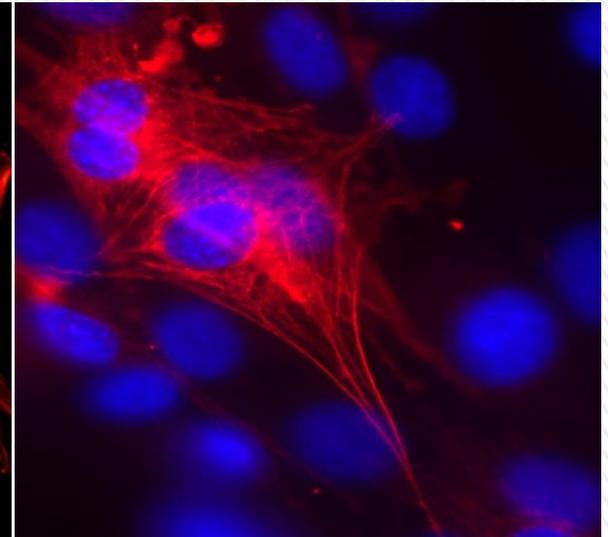
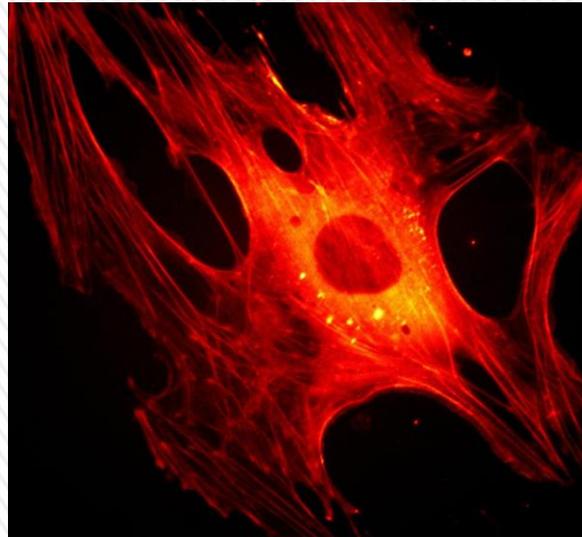
- + Activity reporter plasmid (Blasticidin resistance)
- + targeting construct



selection on Blas (8 days)



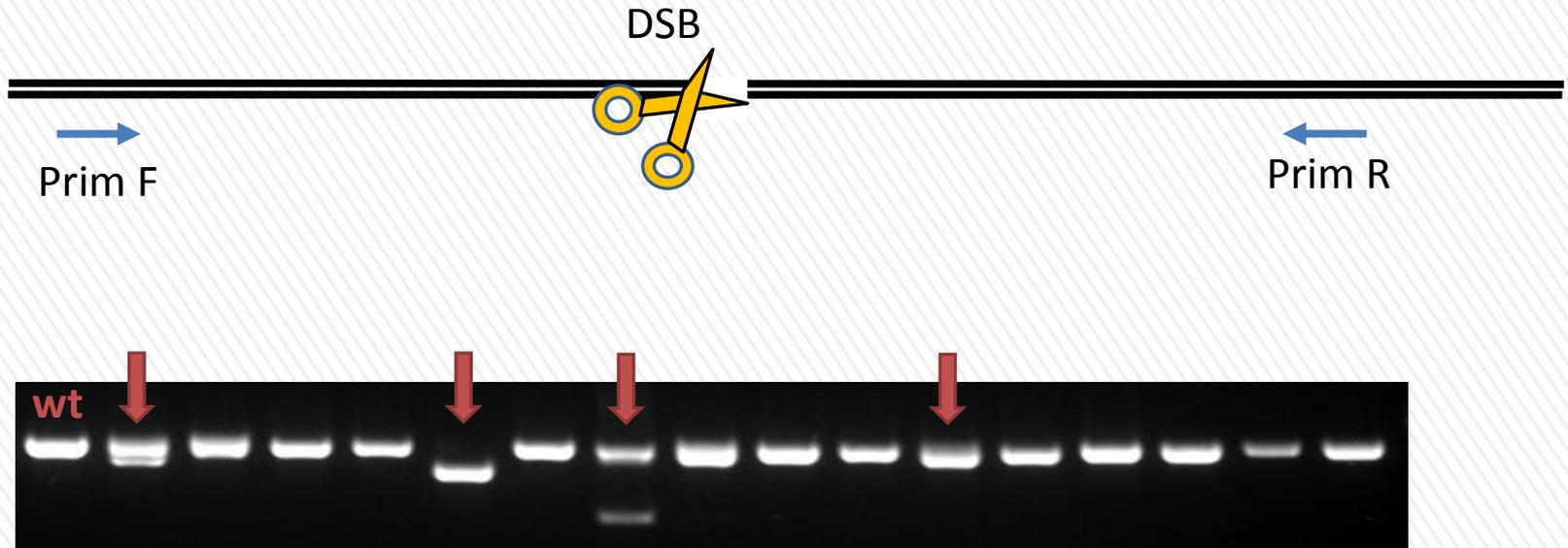
FACS



Kasperek- *unpublished data*

Verification of TALEN activity (PCR)

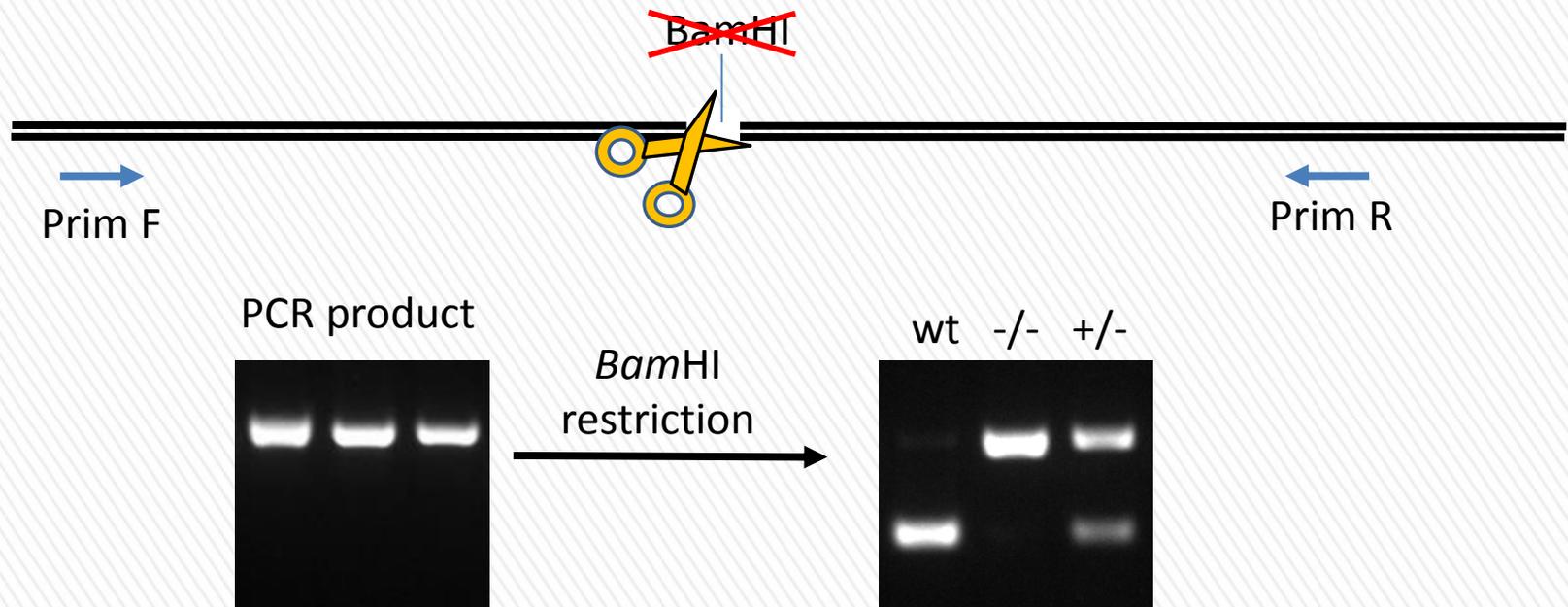
PCR amplification, DNA electrophoresis to analyze variability of PCR products



Verification of TALEN activity (RFLP)

Restriction Fragment Length Polymorphism

- mutation of endogenous restriction site by TALENs
- digestion of PCR product by restriction enzyme



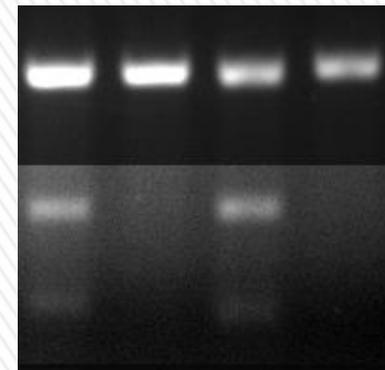
Verification of TALEN activity (T7 endonuclease)

mismatch-specific DNA endonuclease detection of short indels



T7 endonuclease cleavage site

cl1 cl2 cl3 cl4



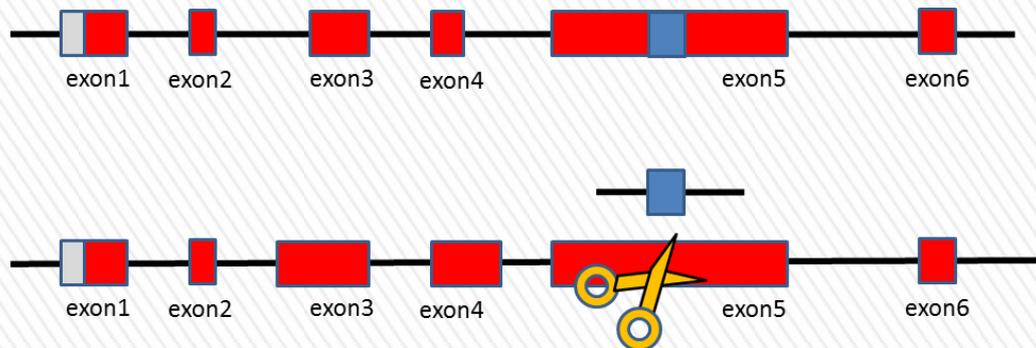
Programmable nucleases:

in vivo model

Examples - Knock-out mutant

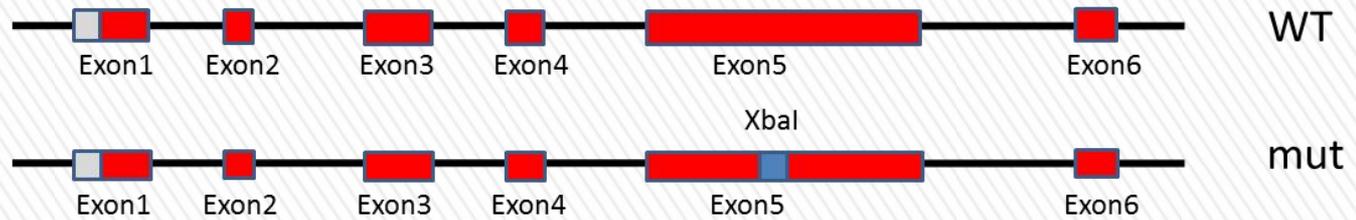
Netherton syndrome

- disorder that affects the skin, hair, and immune system - autosomal recessive (1:200 000)
- chronic skin inflammation in epidermis, abnormal desquamation, disrupted epidermal barrier
- Point mutation in exon 5 Spink 5 gene

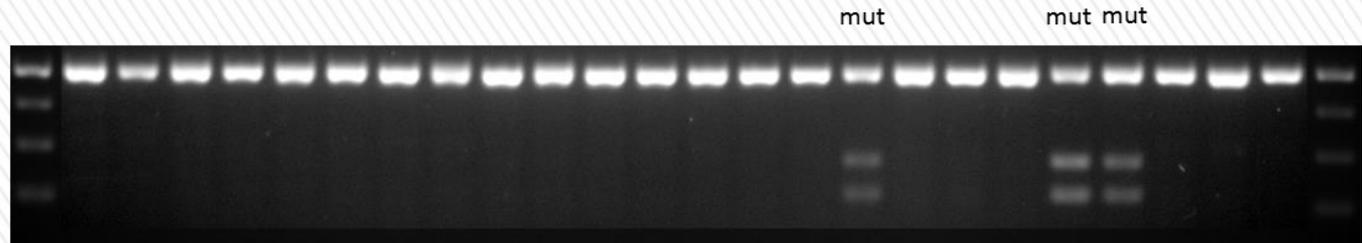


Kasperek- unpublished data

Examples - Knock-out mutant



Spink5 PCR
- Xbal



Kasperek- *unpublished data*

Examples - Knock-out mutant

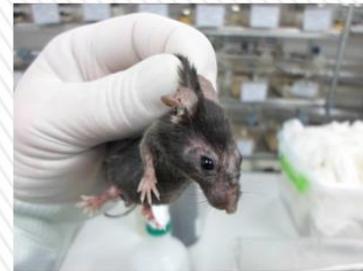
2 days



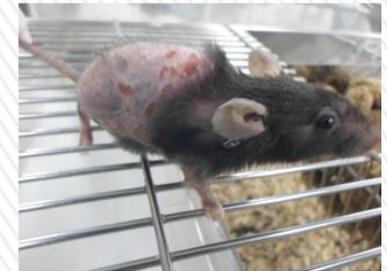
9 days



18 weeks



26 weeks



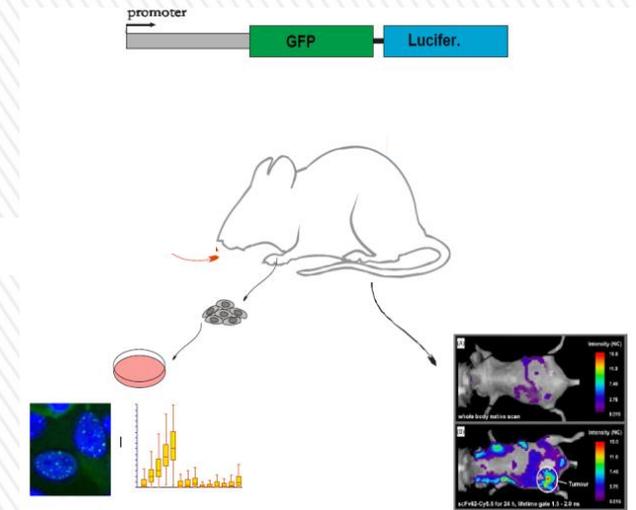
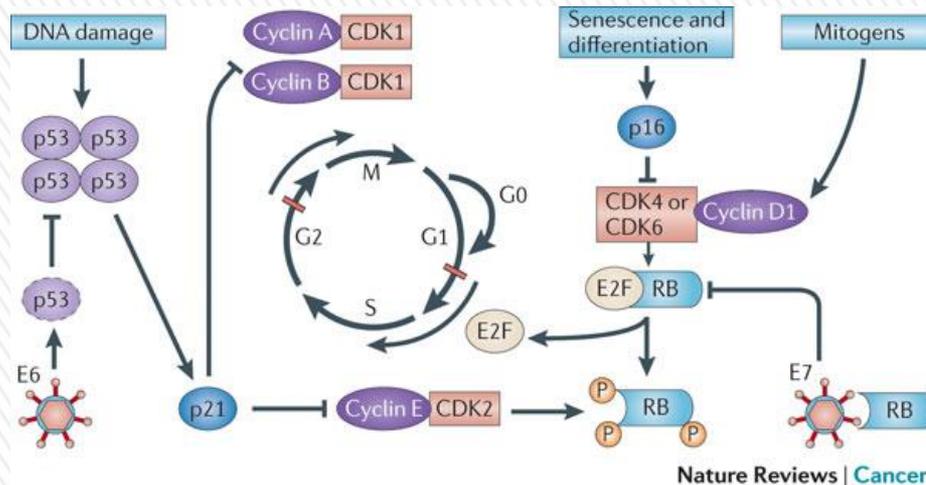
Kasperek- *unpublished data*

Examples - Reporter mice

p21 promoter reporter mice for study of aging

p21^{WAF1/CIP1} - cyclin-dependent kinase inhibitor 1

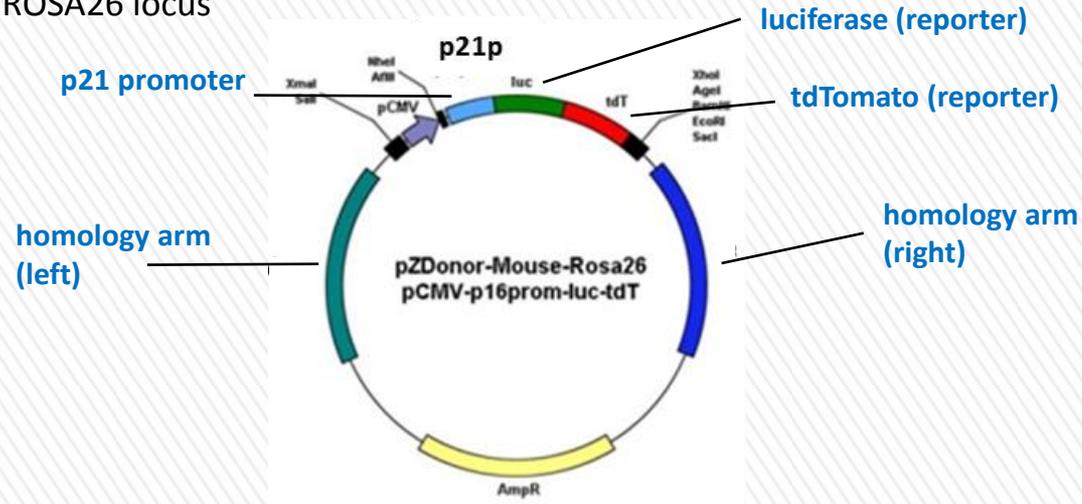
Function : regulator of cell cycle progression at G1 and S phase



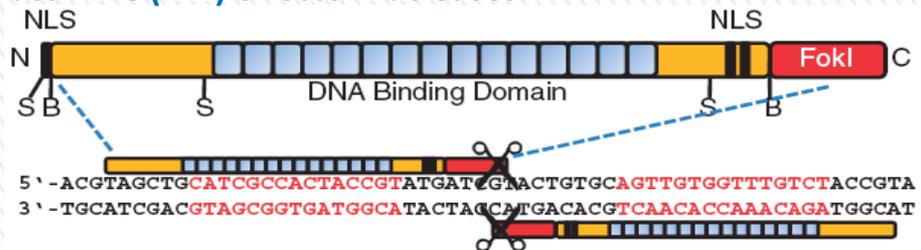
Examples - Reporter mice

Where and how to insert p21 reporter construct?

ROSA26 is a locus used for constitutive, ubiquitous gene expression, over 130 knock-in lines have been created based on the ROSA26 locus

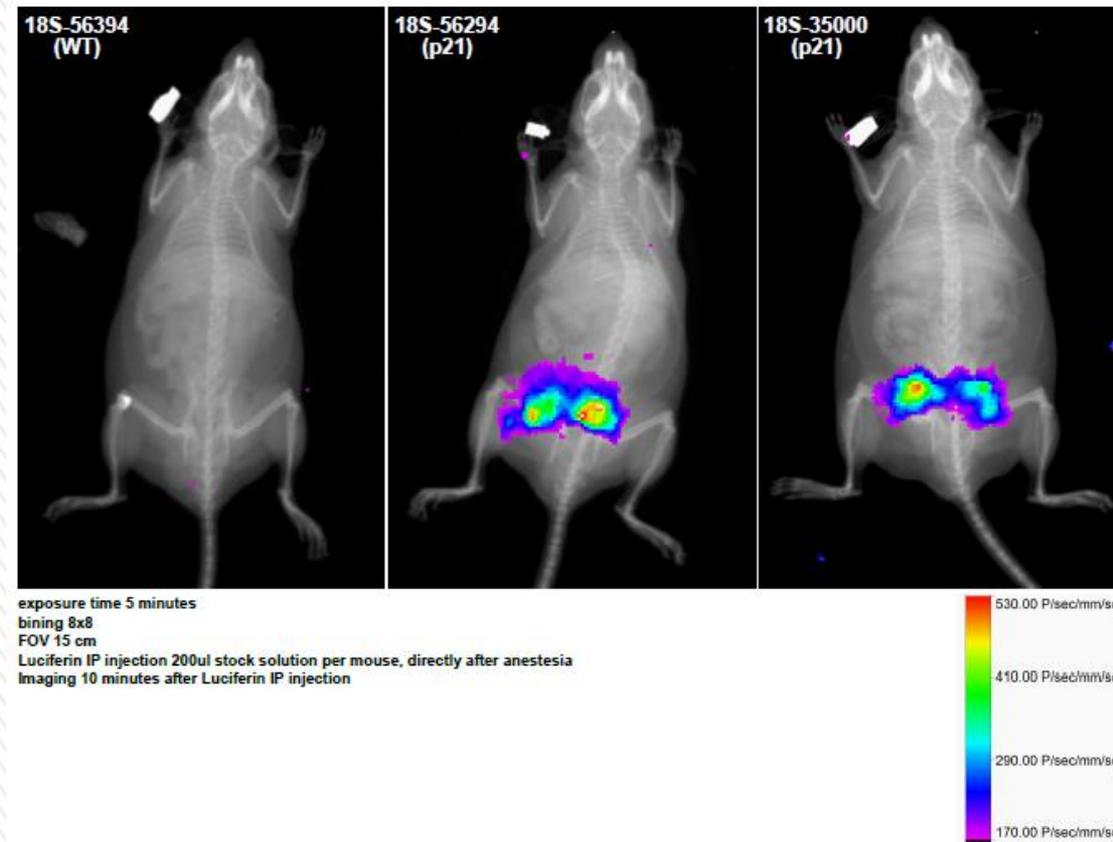


TALENs - transcription activator-like (TAL) effector nucleases



Cermak et al., Nucleic Acids Research, 2011, 1-

Examples- Reporter mice



Conclusions

- both TALEN and CRISPR/Cas9 technology represent very efficient and elegant techniques for gene targeting (knock-out as well as knock-in)
- specific gene knock-out and knock-in is powerful tool for understanding of its role (models of disease)
- possibility of construction of *in vivo* (animals) and *in vitro* (cell culture) models using programmable nucleases



INVESTMENTS IN EDUCATION DEVELOPMENT

Radislav Sedlacek, Assoc. Prof., PhD (*director*)

Phenotyping Module

- Trevor Epp, PhD (*head of module, phenotyping unit*)
- Ivan A. Kanchev, MVSc, DVM (*head, histopathology unit*)
- Karel Chalupsky, PhD (*head, biochemistry unit*)
- Kallayane Chawengsaksohak, PhD (*head, embryogenesis unit*)
- Agnieszka Kubik-Zahorodna, PhD (*head, neurobehavioral unit*)
- Milan Reinis, PhD (*head, immunology unit*)
- Benoit Piavaux, PhD (*head, lung function unit*)
- Jan Polák, MD, PhD (*head, metabolic unit*)
- Jan Procházka, PhD (*head, dysmorphology & whole-body imaging unit*)

Transgenic and Archiving Module

- Inken M. Beck, PhD (*head of module*)
- Irena Jenickova, PhD (*head, ES cell manipulation and transgenesis*)
- Jana Jezkova, MEng.
- Veronika Libova, MSc.
- Irena Placerova, MEng.
- Sandra Potysova, MSc.
- Jana Kopkanova, MEng. (*head, genotyping unit*)
- Dana Kopperova, MSc.
- Monika Volckova, MEng.
- Bjoern Schuster, PhD (*head, targeting unit*)
- Anna Lastuvkova, MSc.
- Henrieta Palesova, MEng.

Animal facility module

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- Peter Neradil, DVM (*deputy head for Vestec animal facility*)
- Marketa Rynekrova (*deputy head for IMG animal facility*)

CCP administrators

- Libor Danek, MA
- Jana Safrankova, BA

OP EC projects

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- | | |
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| Inken M. Beck, PhD | Bjoern Schuster, PhD |
| Monika Cervinkova, PhD | Jolana Tureckova, PhD |
| Trevor Epp, PhD | Libor Danek, MA – project administrator |
| Martin Gregor, PhD | Jana Safrankova, BA – financial administrator |
| Karel Chalupsky, PhD | Radislav Sedlacek, Assoc. Prof., PhD – supervisor |
| Kallyanee Chawengsaksohak, PhD | Ďurí Forejt, Prof., MD, DrSc – supervisor |

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- | | |
|---|------------------------------------|
| Irena Jenickova (née Barnetova), PhD | Iryna Kozmikova, PhD |
| Bohumil Fafilek, PhD | Michaela Krausova, PhD |
| Dominika Fricova, MD, PhD | Silvia Petrezselyova, PhD |
| Zuzana Ileninova, PhD | Vladimir Korinek, PhD – supervisor |
| Miluse Hroudova, PhD | Zbynek Kozmik, PhD – supervisor |
| Slavomir Kinsky, PhD | |
| Jan Kosla, PhD | |
| Radislav Sedlacek, Assoc. Prof., PhD – supervisor | |

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- Chrysoula Pantzartzi, PhD
- Benoit Piavaux, MD, PhD
- Jana Rohozkova, PhD
- Maja Sabol, PhD
- Tomas Venit, PhD
- Pavel Hozak, Prof., PhD – supervisor
- Zbynek Kozmik, PhD – supervisor
- Radislav Sedlacek, Assoc. Prof., PhD – supervisor



1. Which of the following molecular tools DOES NOT belong to genome editing tools?

- a) Zinc finger nucleases b) Tale nucleases **c) RNA ribozymes**

2. What type of direct interaction at the level of DNA is it in case of Crispr/Cas9 system?

- a) protein-DNA **b) RNA-DNA** c) DNA-DNA

3. In which process/feature Cas9 nickase and wild-type Cas9 differ from each other?

- a) wild-type Cas9 creates double stranded DNA breaks, Cas9 nickase creates single stranded DNA breaks**
b) were isolated from different organisms
c) use different cofactors

4. How many of guideRNAs are necessary to create double stranded DNA breaks using the wild-type Cas9?

- a) 1 guideRNA**
b) 2 guideRNA
c) formation of double stranded DNA breaks does not depend on the number of guideRNAs used but on the activity of Cas9

5. What type of DNA repair in cell is used for targeting of a construct into a defined genomic site to generate reporter models?

- a) homologous recombination (HR)**
b) non-homologous end joining (NHEJ)
c) base excision repair (BER)



6. A targeting construct that we plan to use for insertion of a reporter gene into a specific genomic site must contain: 1. reporter gene 2. selection marker and 3.

a) ampicilin

b) homologous arms

c) gen encoding fluorescent protein

7. Which mouse locus is often used for knock-in (insertion of a foreign DNA) strategy?

a) ORF

b) GAL1

c) Rosa26

8. Which of the following genetic elements can be used as reporter gene?

a) promoter

b) gene encoding fluorescent protein or enzyme, the activity of which can be easily measured (e.g. luciferase)

c) gene encoding transcriptional factor

9. Which of the following methods can be used to verify gene insertion into a specific genomic site?

a) luciferase assay

b) PCR, sequencing

c) fluorescence microscopy

10. What is off-target?

a) a process, which leads to „turning off“ of genes that have been modified

b) targeting of several genes by use of programmable nucleases

c) undesired/unplanned/non/specific change in a gene other than that for which programable nuclease have been targeted

